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The importance of the genital tract microflora in the endometriosis development

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Endometriosis is a common pathology among women of reproductive age. In the pathogenesis of its development a great importance is paid to the process of regulation of estrogens and other sex hormones, the implementation of the immune response at the general and local levels, the inflammatory response, and genetic background. In recent years, the role of the microbiota in different parts of the genital tract of the female organism has been studied not only from the standard approach of inflammatory diseases research of the reproductive organs and pathological vaginal discharge, but its contribution into the development of other gynecological diseases, in particular endometriosis, is presented.

The article is devoted to a review of scientific publications about researches devoted to the significance of the genital tract microflora in the pathophysiology of endometriosis. The article presents the concepts of states of eubiosis and dysbiosis, describes the features of the microflora of the lower (vagina, cervix) and upper parts (uterus, fallopian tubes) of the genital tract in healthy women. The data on the importance of Lactobacillus in maintaining an optimal state of acidity in the vaginal environment are presented.

The difference in the composition of the microbiota of the vagina, cervical mucus, uterine cavity and peritoneal fluid in women with and without endometriosis is revealed. The pathogenetic mechanisms of the role of the microbiota in the processes of endometriosis pathogenesis, namely, the regulation of the immune response and inflammation processes in this pathology, are described. It has been shown that in women with endometriosis there are differences in the microflora composition of the vagina, cervix, uterine cavity, peritoneal fluid, manifested by a decreased concentration of Lactobacillus and the presence of a variety of other microorganisms in relation to healthy women. The data about the role of gut microflora in patients with this pathology are also presented. The results of research about the diagnosis of various stages of endometriosis by studying the composition of the genital tract microbiota are described.

Keywords: endometriosis, pathogenesis, vagina, uterus, microbiota, microorganisms, vaginal microflora, microbiome.

Значення мікрофлори генітального тракту у розвитку ендометріозу Л. В. Пахаренко, І. О. Басюга, В. М. Жураківський, О. М. Ласитчук, Н. Я. Курташ

Ендометріоз є поширеною патологією серед жінок репродуктивного віку. У патогенезі його розвитку важливе значення надається процесу регуляції естрогенів та інших статевих гормонів, реалізації імунної відповіді на загальному та локальному рівнях, запальній реакції, генетичному підґрунті. Останніми роками вивчається роль мікробіоти у різних відділах генітального тракту жіночого організму не тільки з позиції вивчення запальних захворювань репродуктивних органів та патологічних вагінальних виділень, але й виникнення та розвитку іншої гінекологічної патології, зокрема ендометріозу.

Стаття присвячена огляду наукових публікацій про значення мікрофлори генітального тракту у патофізіології ендометріозу. У статті наведено поняття станів еубіозу та дисбіозу, описані особливості мікрофлори нижнього (піхва, шийка матки) та верхнього (матка, маткові труби) відділів генітального тракту у здорових жінок. Представлено дані важливості Lactobacillus у підтриманні оптимального стану кислотності у піхвовому середовищі.

Детально розглянуті питання відмінності складу мікробіоти піхви, цервікального слизу, порожнини матки та перитонеальної рідини у жінок з та без ендометріозу. Наведено патогенетичні механізми ролі мікробіоти у процесах патогенезу ендометріозу, а саме — регуляції імунної відповіді та процесів запалення за даної патології. Зазначено, що у жінок з ендометріозом існують відмінності у складі мікрофлори піхви, шийки матки, порожнини матки, перитонеальної рідини, які проявляються зменшенням концентрації Lactobacillus та наявністю різноманіття інших мікроорганізмів щодо здорових жінок. Також представлено дані про роль мікрофлори кишечника у хворих із даною патологією. Описано результати досліджень стосовно діагностики різних стадій ендометріозу за допомогою вивчення складу мікробіоти генітального тракту. Ключові слова: ендометріоз, патогенез, піхва, матка, мікробіота, мікроорганізми, вагінальна мікрофлора, мікробіом.

Microbiota of a woman's genital tract reflects the state of her gynecological and reproductive health [1]. Eubiosis means the state a heathy and balanced ecosystem. In the upper part of the genital tract it is characterized by the presence of >90 % Lactobacillus spp., Bacteroides, Bifidumbacterium Pseudomonas, a low microbial load, the ability to restore tissues, moderate immune stimulation, and in the lower part of the genital tract — a slight diver-

sity $(\alpha-\beta)$ of microflora, the dominance of Lactobacillus which correspondence to I, II, III and V types of microbial communities in the vaginal environment according to J. Ravel et al. [2].

Several types of microorganisms are included into the vaginal microbiota. The dominance of various species of Lactobacillus in the lower part of the genital tract is a sign of the health of the female reproductive system [3].

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In healthy women a small variety of these microorganisms was found in the vagina [2, 4]. Lactobacillus provide stability of the vaginal environment [1, 2, 3, 5, 6]. However, the diversity of the composition of the vaginal microbiome is really large. The results of a metagenomic analysis of the vaginal microbiome of healthy women in the Chinese population, obtained by F. Liu et al., determined 111 types of microorganisms [7].

Lactobacillus contribute to the prevention of genital infections by maintaining optimal acidity in the vaginal environment [8, 9]. This is ensured by the fact that Lactobacillus and other enzymatic microorganisms synthesize lactic acid [6], for the production of which Lactobacillus use glycogen [4, 7]. In addition, a significant number of microorganisms in the vagina, including Lactobacillus, secrete amylase, which breaks down glycogen into simpler compounds that support the vital activity of bacterial populations [6].

Estrogen and progesterone also affect the composition of the vaginal microflora. Estrogens contribute to the accumulation of glycogen through the proliferation of vaginal epithelial cells. Progesterone promotes the lysis of vaginal epithelial cells and the release of glycogen, maintaining the pH of the vagina [10]. The mechanisms of the role of estrogens in vaginal immunity are still unclear [9]. However, scientists have established a connection between the immune system and the vaginal microbiota [9, 11].

There are endo- and exogenous factors that lead to changes in the composition of vaginal microflora – menstrual cycle, pregnancy, age, ethnicity, race, use of contraceptives, sexual activity, etc. [2, 5, 6, 8, 10, 12]. However, the results of B. Chaban et al. research demonstrated that during the menstrual cycle in healthy women the composition of the vaginal microbiota remains relatively stable [13].

A difference in the composition of microbial communities in the cervical canal, uterus, fallopian tubes, and peritoneal fluid compared to the vagina was found [14]. Lactobacillus also predominate in cervical mucus, but their percentage relative to the vagina is less [15]. According to the literature review, it was established that Lactobacillus species predominate, except for the vagina, in endometrium. Greater variability of Lactobacillus is observed in fallopian tubes and ovaries compared to the lower parts of the genital tract [2].

The uterine cavity is not sterile, as it was thought until recently. However, studies indicate that the microbial biomass of the endometrium is 100-10,000 times less than the bacterial content of the vagina [16]. I. Moreno et al. divided the endometrial microbiota into categories: Lactobacillus dominant (LD), in which the content of Lactobacillus spp. is >90%, and non-Lactobacillus dominant (NLD), which is characterized by the proportion of Lactobacillus spp. at the level of <90% and the rate of other bacteria is >10% [17]. The scientists have found that the microbial composition of the uterus differs from the vagina and contains up to 191 operational taxonomic units [17].

Dysbiosis is a non-homeostatic condition, the signs of which in the upper parts of the genital tract are a decrease in the content of Lactobacillus spp. (NLD) to less than 90 %, the detection of Staphylococcus spp., Enterobacter spp., a significant microbial load (infection), tissue dam-

age, immune stimulation [2]. In the lower parts, signs of dysbiosis are a significant diversity of microorganisms, the presence of type IV microbial community (Anaerobic species, Gardnerella, Prevotella, Atopobium) [2].

Bacterial vaginosis and anaerobic polybacterial dysbacteriosis are the most common changes in the vaginal microbiome [18]. Vaginal dysbiosis is known to be associated with increased sensitivity and susceptibility to human immunodeficiency virus, transmission of human immunodeficiency virus and other sexually transmitted microorganisms, an increased risk of development of inflammatory processes of the female genital organs, preterm labor, maternal infections during pregnancy and neonatal infections [18].

It has been established that the gut microbiota affects the intestinal environment, which influence on the distant organs [19]. Thus, changes in the composition of the gut microbiota can contribute to the development of such gynecological pathologies as polycystic ovary syndrome, endometriosis, and cancer [19, 20].

Endometriosis is a hormonally depended gynecological pathology, which is diagnosed in approximately 10 % of women in reproductive age. The reason of the pathology is the growth of the endometrium outside the uterine cavity [21]. The main symptoms of endometriosis are chronic pelvic pain, infertility, menstrual cycle disorders, and so on [22].

It was found that changes in estrogen metabolism lead not only to the development of endometriosis, but also they contribute to inflammation of the endometrium, changes in the vaginal microbiota, and genetic factors play an important role in the pathogenesis of the disease [23].

Researchers believe that the NLD microbiota is not only associated with adverse pregnancy outcomes or bacterial vaginosis, but can provoke an inflammatory response in the endometrium, which is one of the main factors in the development of endometriosis, so it should be considered that the changed endometrial microbiota or the dominance of NLD is related to this pathology [17].

The mechanisms of endometriosis, namely, disorders in the immunoregulatory functions of the microbiota due to dysbacteriosis have a lot of common signs with such diseases as inflammatory bowel disease, psoriasis, arthritis, and cancer [24]. It is known that dysbiosis in the intestines and vagina leads to changes in the functioning of the immune system, namely, to an increase of pro-inflammatory cytokines, damage of immune cells, which plays a role in the pathogenesis of endometriosis [24]. Immune dysregulation can subsequently contribute to the appearance of a chronic inflammatory process, which together with the activization of mechanisms of adhesion and angiogenesis, leads to the progression of endometriosis [24].

One of the explanations for the occurrence of endometriosis is the theory of the retrograde menstrual blood flow through the fallopian tubes to the abdominal cavity. The difference in the immune response in the peritoneal environment may be a hypothesis why in some cases endometrioid lesions appear, in particular, on the intestine and peritoneum [25, 26, 27, 28]. A more pronounced immune response will be in the cases of the intestinal dysbacteriosis, because intestinal bacteria stimulate neutrophils, acti-

vate CD4 T-cells, while the barrier function is disturbed, the release of microbial metabolites occurs, and this leads to inflammatory changes, an increase in the number of peritoneal macrophages, the ability of which is somewhat limited in phagocytosis of new implanted endometrial foci, thus a long-term inflammatory process develops, the persistence of foci and progression of endometriosis occurs [25, 26, 29, 30].

A study of W. Wei et al. indicates that there is a difference in the composition of the microbiota in different parts of the reproductive tract in women with endometriosis. In the lower parts, in particular, in the lower third of the vagina and the posterior vault, Lactobacillus predominate. Starting from the cervix and in the upper parts of the genital tract (endometrium, peritoneal fluid), a more pronounced diversity in the composition of the microbiota was found, namely, an increase in the number of operational taxonomic units, which may indicate the presence of a bacterial component in the development of the disease [31].

The association between the bacterial factor and endometriosis is emphasized by the studies of the groups of scientists W. C. Lin et al. and F. W. Tai et al., which demonstrated that women with infectious processes of the lower part of the genital tract or inflammatory processes of the internal genital organs in the anamnesis have an increased risk of endometriosis in 2-3 times [32, 33]. The results of the study of J. Kunaseth et al. confirm a similar proportion of Lactobacillus in the composition of the microbiota of the vaginal environment in women with adenomyosis (51.20 %) and women without adenomyosis (50.54 %) [34]. However, scientists have found an increased number of operational taxonomic units of vaginal microorganisms such as Alloscardovia, Oscillospirales, Ruminoccoccaceae, UCG 002, Oscillospiraceae, Enhydrobacter, Megamonas, Moraxellaceae, Subdoligranulum, Selenomonadaceae and Faecalibacterium in patients with adenomyosis [34].

The study of the vaginal microbiota for the determination the endometriosis stage according to the updated recommendations of the American Society for Reproductive Medicine (revised American Society for Reproductive Medicine) is being considered. It was found that the vaginal and gut microbiota is associated with the degree of endometriosis and the presence of 1-2 stages of endometriosis can be most accurately assumed than 3-4 stages. At the same time, the operational anaerobic unit from the genus Anaerococcus was found to be the greatest prognostic criterion for establishing the stages of the disease [35].

It was found that women with endometriosis/adenomyosis and clinical manifestations of chronic pelvic pain had an increased number of Clostridium butyricum, Clostridium disporicum, Alloscardovia omnicolens, Veillonella montpellierensis in the vaginal environment compared to women with chronic pelvic pain without endometriosis/adenomyosis and women without chronic pelvic pain [36]. The presence of such microorganisms as Clostridium butyricum, Clostridium disporicum, Alloscardovia omnicolens and Veillonella montpellierensis in the vaginal environment, as well as the simultaneous deficiency of Lactobacillus jensenii, Lactobacillus reuteri and Lactobacillus iners can be considered as potential pathogens and markers

of chronic pelvic pain that occurs in women with the background of endometriosis/adenomyosis. When the relative amount of Clostridium disporicum exceeds 0.001105 % and Lactobacillus reuteri is less than 0.1911349 %, the differential diagnostic sensitivity and specificity are 81.08 % and 52.0 %, respectively. When CA125 in blood serum is included in the diagnostic algorithm, the sensitivity of the method increases to 89.19 %, but the specificity remains unchanged at the level of 52.0 % [36].

In women with endometriosis there is a tendency to the increased concentration of Firmicutes pathogens and a decreased in the content of Actinobacteria and Bacteroides bacteria in the cervix. In the late stages of endometriosis, deep infiltrative endometriosis, the presence of a high level of CA125, as well as in patients with a pronounced pain syndrome and infertility caused by endometriosis, there is an unstable content of Lactobacillus in the cervical canal in combination with an increase in the level of Streptococci and a decrease in the content of microorganisms of the Dialister group [37]. The study of K. Akiyama et al. presents that although Lactobacillus spp. prevailed in the cervical mucus in women with endometriosis, there was an increased content of such pathogens as Corynebacterium, Enterobacteriaceae, Flavobacterium, Pseudomonas and Streptococcus in the patients. Of these microorganisms, the highest concentration was determined for Enterobacteriaceae and Streptococcus [38]. A group of scientists head by B. Ata established that in the cervical environment in patients with III-IV stages of endometriosis there is an increased content of pathogenic bacteria (Gardnerella, Shigella, Streptococcus, Escherichia, Ureaplasma) [39].

In patients with endometriosis compared to healthy women there are changes in endometrial microbiota composition. Thus, there is a decrease in the concentration of Lactobacilli in the endometrium and an increase in the number of pathogens that contribute to the development of bacterial vaginosis and the high concentration of opportunistic bacteria are determined in the patients with endometriosis [24]. In women with endometriosis among the bacterial taxa in the endometrium microbiota grampositive (Actinobacteria phylum, Streptococcaceae) and gram-negative microorganisms (Oxalobacteria, Tepidimonas) are often detected, while in patients with a pain syndrome that is not related to endometriosis, which was confirmed during surgery interventions, and persons with other benign gynecological pathology, often gram-negative pathogens (group Burkholderiaceae and Ralstonia) are established [40]. The endometrial microbiota in endometriosis is enriched by Actinobacteria phylum, Oxalobacteraceae and Streptococcaceae families, and Tepidomonas genus relative to symptomatic patients without endometriosis [40].

In the women with endometriosis in the peritoneal fluid according to the results of W. Yaun et al. study the slight changes in the microbiota in relation to such pathogens as Acidovorax, Devosia, Methylobacterium, Phascolarctobacterium, Streptococcus was found [41], and research of S-R. Lee et al. indicates an increased content of Acinetobacter, Pseudomonas, Streptococcus, and Enchydrobacter and a reduced number of Propionibacterium, Actinomyces, and Rothia relative to control women [42].

However, scientific data regarding the importance of the microbiota of the genital tract in the genesis of endometriosis are controversial. According to the research of S. Hernandes et al. a similar composition of microorganisms in the vaginal environment, eutopic endometrium and endometrioid foci, consisting of Lactobacillus, Gardnerella, Streptococcus and Prevotella, was found between healthy women and patients with endometriosis. However, deep endometriosis foci have been detected to have a dissimilar bacterial composition with a lower number of Lactobacillus and a higher number of Alishewanella, Enterococcus, and Pseudomonas [43].

More and more studies are finding a relation between gut microbiota and endometriosis. Gut microflora can play a role in the mechanisms of endometriosis development through the processes of estrogen regulation and immune inflammation [26, 44, 45]. The studies on the animals suggest a two-way relationship between the gut microbiota and the onset and progression of endometriosis [23]. It was established that the assessment of the state of the gut microbiota is more informative for the diagnosis of endometriosis than the composition of the microbiota of the cervix [46].

Thus, it remains important to study the role of the genital tract microbiota in the development of endometriosis, which is confirmed by many researches.

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REFERENCES

- 1. Zhu B, Tao Z, Edupuganti L, Serrano MG, Buck GA. Roles of the Microbiota of the Female Reproductive Tract in Gynecological and Reproductive Health. Microbiol Mol Biol Rev. 2022;86(4):e0018121. doi: 10.1128/mmbr.00181-21.
- 2. Punzón-Jiménez P, Labarta E. The impact of the female genital tract microbiome in women health and reproduction: a review. J Assist Reprod Genet. 2021;38(10):2519-41. doi: 10.1007/s10815-021-02247-5.
- 3. Nunn KL, Clair GC, Adkins JN, Engbrecht K, Fillmore T, Forney LJ. Amylases in the Human Vagina. mSphere. 2020;5(6):e00943-20. doi: 10.1128/mSphere.00943-20.
- 4. Cheng L, Gao Y, Xia Q, Wang H, Xie X, Liu Y, et al. Reproductive tract microbiota of women in childbearing age shifts upon gynecological infections and menstrual cycle. BMC Microbiol. 2021;21(1):252. doi: 10.1186/s12866-021-02300-4.
- 5. Saraf VS, Sheikh SA, Ahmad A, Gillevet PM, Bokhari H, Javed S. Vaginal microbiome: normalcy vs dysbiosis. Arch Microbiol. 2021;203(7):3793-802. doi: 10.1007/s00203-021-02414-3.
- 6. Buchta V. Vaginal microbiome. Ceska Gynekol. 2018;83(5):371-9.
- 7. Liu F, Zhou Y, Zhu L, Wang Z, Ma L, He Y, Fu P. Comparative metagenomic analysis of the vaginal microbiome in healthy women. Synth Syst Biotechnol.

- 2021;6(2):77-84. doi: 10.1016/j.syn-bio.2021.04.002.
- 8. Chopra C, Bhushan I, Mehta M, Koushal T, Gupta A, Sharma S, Kumar M, et al. Vaginal microbiome: considerations for reproductive health. Future Microbiol. 2022;17:1501-13. doi: 10.2217/fmb-2022-0112.
- 9. Villa P, Cipolla C, D'Ippolito S, Amar ID, Shachor M, Ingravalle F, Scaldaferri F, Puca P, Di Simone N, Scambia G. The interplay between immune system and microbiota in gynecological diseases: a narrative review. Eur Rev Med Pharmacol Sci. 2020;24(10):5676-90. doi: 10.26355/eurrev 202005 21359.
- 10. Shen L, Zhang W, Yuan Y, Zhu W, Shang A. Vaginal microecological char-

- acteristics of women in different physiological and pathological period. Front Cell Infect Microbiol. 2022;12:959793. doi: 10.3389/fcimb.2022.959793.
- 11. Gholiof M, Adamson-De Luca E, Wessels JM. The female reproductive tract microbiotas, inflammation, and gynecological conditions. Front Reprod Health. 2022;4:963752. doi: 10.3389/frph.2022.963752.
- 12. Pramanick R, Parab S, Mayadeo N, Warke H, Aranha C. Cross sectional analysis of vaginal Lactobacillus in asymptomatic women of reproductive age in Mumbai, India. J Infect Dev Ctries. 2018;12(12):1096-104. doi: 10.3855/jidc.10154.

- 13. Chaban B, Links MG, Jayaprakash TP, Wagner EC, Bourque DK, Lohn Z, et al. Characterization of the vaginal microbiota of healthy Canadian women through the menstrual cycle. Microbiome. 2014;2:23. doi: 10.1186/2049-2618-2-23.
- 14. Chen C, Song X, Wei W, Zhong H, Dai J, Lan Z, Li F, et al. The microbiota continuum along the female reproductive tract and its relation to uterine-related diseases. Nat Commun. 2017;8(1):875. doi: 10.1038/s41467-017-00901-0.
- 15. Chen S, Gu Z, ZhangW, Jia S, Wu Y, Zheng P, et al. Microbiome of the lower genital tract in Chinese women with endometriosis by 16s-rRNA sequencing technique: a pilot study. Ann Transl Med. 2020;8:1440. doi: 10.21037/atm-20-1309.
- 16. Baker JM, Chase DM, Herbst-Kralovetz MM. Uterine Microbiota: residents, tourists, or invaders? Front Immunol. 2018;9:1-16. doi: 10.3389/fimmu.2018.00208.
- 17. Moreno I, Codoñer FM, Vilella F, Valbuena D, Martinez-Blanch JF, Jimenez-Almazán J, et al. Evidence that the endometrial microbiota has an effect on implantation success or failure. Am J Obstet Gynecol. 2016;215(6):684-703. doi: 10.1016/j.ajog.2016.09.075.
- 18. van de Wijgert JHHM, Jespers V. The global health impact of vaginal dysbiosis. Res Microbiol. 2017;168(9-10):859-64. doi: 10.1016/i.resmic.2017.02.003.
- 19. Qi X, Yun C, Pang Y, Qiao J. The impact of the gut microbiota on the reproductive and metabolic endocrine system. Gut Microbes. 2021;13(1):1-21. doi: 10.1080/19490976.2021.1894070.
- 20. Svensson A, Brunkwall L, Roth B, Orho-Melander M, Ohlsson B. Associations Between Endometriosis and Gut Microbiota. Reprod Sci. 2021;28(8):2367-77. doi: 10.1007/s43032-021-00506-5.
- 21. World Health Organisation. Endometriosis [Internet]. Geneva: WHO; 2021. Available from: https://www.who.int/news-room/fact-sheets/detail/endo-

- metriosis #:~:text= Endometriosis%20 is%20a%20disease%20where,and%20 qirls%20qloballv%20(2).
- 22. Gruber TM, Mechsner S. Pathogenesis of Endometriosis: The Origin of Pain and Subfertility. Cells. 2021;10(6):1381. doi: 10.3390/cells10061381.
- 23. Salliss ME, Farland LV, Mahnert ND, Herbst-Kralovetz MM. The role of gut and genital microbiota and the estrobolome in endometriosis, infertility and chronic pelvic pain. Hum Reprod Update. 2021;28(1):92-131. doi: 10.1093/humupd/dmab035.
- 24. Jiang I, Yong PJ, Allaire C, Bedaiwy MA. Intricate Connections between the Microbiota and Endometriosis. Int J Mol Sci. 2021;22(11):5644. doi: 10.3390/ijms22115644.
- 25. Zondervan KT, Becker CM, Koga K, Missmer SA, Taylor RN, Viganò P. Endometriosis. Nat Rev Dis Primers. 2018;4(1):9. doi: 10.1038/s41572-018-0008-5.
- 26. Laschke MW, Menger MD. The gut microbiota: a puppet master in the pathogenesis of endometriosis? Am J Obstet Gynecol. 2016;215(1):68.e1-4. doi: 10.1016/j.ajog.2016.02.036.
- 27. Symons LK, Miller JE, Kay VR, Marks RM, Liblik K, Koti M, et al. The Immunopathophysiology of Endometriosis. Trends Mol Med. 2018;24(9):748-62. doi: 10.1016/j.molmed.2018.07.004.
- 28. Nothnick WB. Treating endometriosis as an autoimmune disease. Fertil Steril. 2001;76(2):223-31. doi: 10.1016/s0015-0282(01)01878-7.
- 29. Ivanov II, Atarashi K, Manel N, Brodie EL, Shima T, Karaoz U, et al. Induction of intestinal Th17 cells by segmented filamentous bacteria. Cell. 2009;139(3):485-98. doi: 10.1016/j.cell.2009.09.033.
- 30. Yuan M, Li D, Zhang Z, Sun H, An M, Wang G. Endometriosis induces gut microbiota alterations in mice. Hum Reprod. 2018;33(4):607-16. doi: 10.1093/humrep/dex372.
- 31. Wei W, Zhang X, Tang H, Zeng L, Wu R. Microbiota composition and distribu-

- tion along the female reproductive tract of women with endometriosis. Ann Clin Microbiol Antimicrob. 2020;19(1):15. doi: 10.1186/s12941-020-00356-0.
- 32. Lin WC, Chang CY, Hsu YA, Chiang JH, Wan L. Increased Risk of Endometriosis in Patients With Lower Genital Tract Infection: A Nationwide Cohort Study. Medicine (Baltimore). 2016;95(10):e2773. doi: 10.1097/MD.0000000000002773.
- 33. Tai FW, Chang CY, Chiang JH, Lin WC, Wan L. Association of Pelvic Inflammatory Disease with Risk of Endometriosis: A Nationwide Cohort Study Involving 141,460 Individuals. J Clin Med. 2018;7(11):379. doi: 10.3390/jcm7110379.
- 34. Kunaseth J, Waiyaput W, Chanchaem P, Sawaswong V, Permpech R, Payungporn S, et al. Vaginal microbiome of women with adenomyosis: A case-control study. PLoS One. 2022;17(2):e0263283. doi: 10.1371/journal.pone.0263283.
- 35. Perrotta AR, Borrelli GM, Martins CO, Kallas EG, Sanabani SS, Griffith LG, et al. The Vaginal Microbiome as a Tool to Predict rASRM Stage of Disease in Endometriosis: a Pilot Study. Reprod Sci. 2020;27(4):1064-73. doi: 10.1007/s43032-019-00113-5.
- 36. Chao X, Liu Y, Fan Q, Shi H, Wang S, Lang J. The role of the vaginal microbiome in distinguishing female chronic pelvic pain caused by endometriosis/adenomyosis. Ann Transl Med. 2021;9(9):771. doi: 10.21037/atm-20-4586.
- 37. Chang CY, Chiang AJ, Lai MT, Yan MJ, Tseng CC, Lo LC, et al. A More Diverse Cervical Microbiome Associates with Better Clinical Outcomes in Patients with Endometriosis: A Pilot Study. Biomedicines. 2022;10(1):174. doi: 10.3390/biomedicines10010174.
- 38. Akiyama K, Nishioka K, Khan KN, Tanaka Y, Mori T, Nakaya T, et al. Molecular detection of microbial colonization in cervical mucus of women with and without endometriosis. Am J Reprod Immunol. 2019;82(2):e13147. doi: 10.1111/aji.13147.

- 39. Ata B, Yildiz S, Turkgeldi E, Brocal VP, Dinleyici EC, Moya A, et al. The Endobiota Study: Comparison of Vaginal, Cervical and Gut Microbiota Between Women with Stage 3/4 Endometriosis and Healthy Controls. Sci Rep. 2019;9(1):2204. doi: 10.1038/s41598-019-39700-6.
- 40. Wessels JM, Domínguez MA, Leyland NA, Agarwal SK, Foster WG. Endometrial microbiota is more diverse in people with endometriosis than symptomatic controls. Sci Rep. 2021;11(1):18877. doi: 10.1038/s41598-021-98380-3.
- 41. Yuan W, Wu Y, Chai X, Wu X. The colonized microbiota composition in the peritoneal fluid in women with endometriosis. Arch Gynecol Obstet. 2022;305(6):1573-80. doi: 10.1007/s00404-021-06338-7.
- 42. Lee SR, Lee JC, Kim SH, Oh YS, Chae HD, Seo H, et al. Altered Composition of Microbiota in Women with Ovarian Endometrioma: Microbiome Analyses of Extracellular Vesicles in the Peritoneal Fluid. Int J Mol Sci. 2021;22(9):4608. doi: 10.3390/ijms22094608.
- 43. Hernandes C, Silveira P, Rodrigues Sereia AF, Christoff AP, Mendes H, et al. Microbiome Profile of Deep Endometriosis Patients: Comparison of Vaginal Fluid, Endometrium and Lesion. Diagnostics (Basel). 2020;10(3):163. doi: 10.3390/diagnostics10030163.
- 44. Qin R, Tian G, Liu J, Cao L. The gut microbiota and endometriosis: From pathogenesis to diagnosis and treatment. Front Cell Infect Microbiol. 2022;12:1069557. doi: 10.3389/fcimb.2022.1069557.
- 45. Talwar C, Singh V, Kommagani R. The gut microbiota: a double-edged sword in endometriosis†. Biol Reprod. 2022;107(4):881-901. doi: 10.1093/biolre/ioac147.
- 46. Huang L, Liu B, Liu Z, Feng W, Liu M, Wang Y, et al. Gut Microbiota Exceeds Cervical Microbiota for Early Diagnosis of Endometriosis. Front Cell Infect Microbiol. 2021;11:788836. doi: 10.3389/fcimb.2021.788836.

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